Article ID: 89311 DOI: 10.5586/asbp.89311

#### Publication History Received: 2020-05-01 Accepted: 2020-09-02 Published: 2020-11-12

#### Handling Editor

Agnieszka Popiela, University of Szczecin, Poland; https://orcid.org/0000-0001-9297-0538

#### Authors' Contributions

MKu prepared the core parts of introduction, first second, third, and sixth chapters, and conclusions; MKu, MKo, and BGK prepared the fourth chapter; BGK prepared fifth chapter; MKo provided data on the photobionts of Graphidaceae; all authors read and corrected the entire text

#### Funding

This research received support from the National Science Center (project No. 2015/17/B/NZ8/02441).

#### **Competing Interests**

No competing interests have been declared.

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#### **REVIEW in RECENT DEVELOPMENTS IN TAXONOMY AND PHYLOGENY OF PLANTS**

# One Name – One Fungus: The Influence of Photosynthetic Partners on the Taxonomy and Systematics of Lichenized Fungi

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### Abstract

Lichens are fungi (mycobionts) that form symbiotic associations with photoautotrophic prokaryotes or eukaryotes (photobionts); however, some species can exchange photosynthetic partners during their lifecycles. This phenomenon modifies the morphology of lichens and consequently influences the taxonomy of lichenized fungi. Here, a few such cases in which the photobionts influenced the taxonomy and systematics of lichenized fungi are reviewed. Two different morphotypes of the same species – known as photomorphs – were classified as different species and sometimes different genera. Moreover, different types of photobionts and the absence or presence (optional lichenization) of an alga in the thallus were believed to be diagnostic characters for discriminating genera. However, the taxonomy and systematics of lichens are based always, according to Article F.1.1. of the *International Code of Nomenclature for Algae, Fungi, and Plants*, on the fungal partner and only one name is applied.

#### Keywords

mycobiont; photobiont; photomorphs; optional lichenization; Hymeneliaceae; Lecanographaceae; Peltigeraceae; Stictidaceae

#### 1. Introduction

Lichens, also known as lichenized fungi, are an iconic example of symbiosis and belong to Ascomycota Caval.-Sm. or Basidiomycota R. T. Moore (Lücking, Hodkinson, & Leavitt, 2017). Their thalli consist mainly of the hyphae of the lichenized fungus (called mycobiont in symbiosis) as well as various heterotrophic and autotrophic eukaryotic and prokaryotic organisms. The mycobiont is associated with one or more photosynthetic partners (photobionts), which may belong to eukaryotic green algae or prokaryotic cyanobacteria (e.g., Dal Grande et al., 2017; Friedl & Büdel, 2008; Lücking et al., 2009; Moya et al., 2017; Onuț-Brännström et al., 2018; Purvis, 2000). Lichen thalli also harbor other fungi, and this mycobiome is formed by the following three groups: (i) symptomatic lichenicolous fungi (i.e., showing their reproductive structures, which distinguishes them from endolichenic fungi), (ii) asymptomatic endolichenic fungi, and (iii) extraneous fungi and their diaspores (e.g., Arnold et al., 2009; Bates et al., 2012; Diederich et al., 2018; Fernández-Mendoza et al., 2017; Flakus et al., 2019; Muggia et al., 2016; Spribille et al., 2016). The third group probably does not play any ecological role in lichen symbiosis; however, these lichen thalli may serve as banks of spores and mycelia or reservoirs of the local fungal mycobiome (Fernández-Mendoza et al., 2017). Lichen thalli also contain nonphotosynthetic bacteria, which contribute to the functioning of the symbiotic system of the lichen as a whole (e.g., Bates et al., 2011, 2012; Grube et al., 2015; West et al., 2017).

The most common lichen photobionts belong to Chlorophyta Reichenbach, and nearly 90% of lichenized fungi are associated with green algae (chlorobionts), mostly of the genera Asterochloris Tschermak-Woess, Trebouxia Puymaly, and Trentepholia Mart. s. l. (e.g., Dal Grande et al., 2017; Friedl & Büdel, 2008; Kosecka et al., 2020; Miadlikowska et al., 2006, 2018; Moya et al., 2017; Onut-Brännström et al., 2018; Rivas Plata et al., 2010; Singh et al., 2019). Some lichenized fungi form thalli with other autotrophic groups of organisms, of which cyanobacteria (cyanobionts) are the commonest symbiotic partners (e.g., Lücking et al., 2009, 2017; Miadlikowska et al., 2006, 2018; Tschermak-Woess, 1988, 1989). In some lichens, both types of photobionts can be present in a single thallus, forming a tripartite association, with green algae being the dominant component of the thallus and cyanobionts recruited in cephalodia (e.g., Lamb, 1951; Miadlikowska et al., 2018; Oset & Kukwa, 2012; Schneider et al., 2016; Tønsberg & Goward, 2001). In a few genera, a single species of lichenized fungus can form two different thalli (photomorphs), with either a chlorobiont (chloromorph) or a cyanobiont (cyanomorph), and this type of association is related to environmental factors (Green et al., 1993; Moncada, Coca, & Lücking, 2013; Purvis, 2000; Ranft et al., 2018; Tønsberg & Goward, 2001). More recently, it was reported that a single thallus of one lichen can associate with numerous species or operational taxonomic units (OTUs) of closely related photobionts and that one or two strains are dominant within the thallus, with the rest of the pool representing associated photobionts (e.g., del Campo et al., 2013; Dal Grande et al., 2017; Moya et al., 2017; Onut-Brännström et al., 2018).

In general, the morphology of the lichen thallus is shaped by the mycobiont, but in a few genera of filamentous lichens (e.g., Cystocoleus Thwaites or Cyphellostereum D. A. Reid), the thallus structure depends upon the photobiont filaments and the fungal hyphae surrounding them (e.g., Dal-Forno et al., 2013; Hawksworth et al., 2011). However, in some lichen groups with nonfilamentous photobionts, the morphology of the thallus depends upon autotrophic partners, but in a different way. In other words, the switch from one type of photobiont to another changes the morphological traits (anatomic structure, color, and size of the thallus) or even propagation modes (asexual versus sexual reproduction). Sometimes, in such cases, two names were applied to different morphotypes of the same species (e.g., Ertz et al., 2018; Heidmarsson et al., 1997; Jørgensen, 1998; Miadlikowska et al., 2018; Moncada, Coca, & Lücking, 2013; Tønsberg & Goward, 2001). Nevertheless, the taxonomy and systematics of lichens are based always, according to Article F.1.1. of the International Code of Nomenclature for Algae, Fungi, and Plants, on the fungal partner, and only one name can be applied (Turland et al., 2018). In this review, some examples of the influence of shift in photosynthetic partners on the taxonomy of lichenized fungi are presented.

#### 2. Sticta – A Case of Fishy Lichens

Sticta (Schreb.) Ach. is a genus of large, foliose (rarely fruticose) lichens developing cyphellae (structures on the lower side of thalli that allow gas exchange) – known only in this genus (Moncada, Lücking, & Betancourt-Macuase, 2013; Purvis, 2000). The genus is widely distributed but is more diverse in the tropics and southern hemisphere (e.g., Galloway, 1994, 1997; Lücking, Hodkinson, & Leavitt, 2017; Moncada, Lücking, & Betancourt-Macuase, 2013; Ranft et al., 2018). Similar to many members of Peltigeraceae Dumort., *Sticta* species can associate either with green algae or cyanobacteria. Moreover, similar to that in *Peltigera* Willd. or *Ricasolia* De Not., in some *Sticta* species, the cyanobiont is enclosed in cephalodia or two different photomorphs (chloromorph and cyanomorph; Figure 1A) develop (e.g., Galloway, 1994, 1997, 1998, 2001; Miadlikowska et al., 2018; Moncada, Coca, & Lücking, 2013; Moncada, Lücking, & Betancourt-Macuase, 2013; Ranft et al., 2018; Tønsberg & Goward, 2001). Thalli with cyanobionts also have a very characteristic fish-like smell caused by the production of soluble amines (Galloway & Nash, 2004; James & Henssen, 1976).

The taxonomy of *Sticta* is based on numerous morphological characteristics (e.g., type of thallus, color of the lower surface, and development of the tomentum); however, the type of photobiont was typically the first characteristic in keys dividing

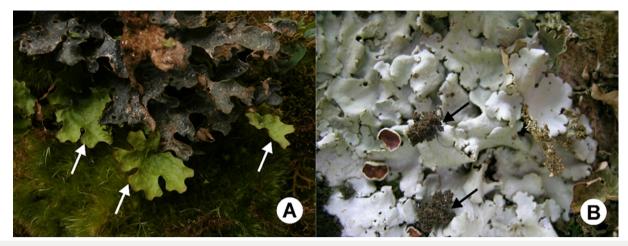


Figure 1 (A) Two photomorphs of *Stica* on the same thallus with chloromorph (arrow) growing out of cyanomorph. (B) *Dendriscocaulon*-like cephalodia (arrow) on thallus of *Ricasolia amplissima* in Scotland.

species into groups (Galloway, 1994, 1997, 1998). This morphology-based taxonomy has obscured the diversity and identity of species, with photomorphs recognized as separate taxa and some of them even placed in different genus, *Dendriscocaulon* Nyl. (e.g., Galloway, 2001; Magain et al., 2012; Moncada, Coca, & Lücking, 2013; Purvis, 2000; Ranft et al., 2018; Tønsberg & Goward, 2001; see also below).

One of the first discoveries that the same *Sticta* species can recruit either a green alga or a cyanobacterium was based on morphological and ecological studies. It was found that environmental factors (light and humidity) are crucial to the selection of photobiont type for thallus development. In more humid and sheltered habitats with low illumination, a cyanobiont is preferred over a chlorobiont, which is the photosynthetic component of the thallus in more dry and open habitats. In intermediate conditions, composite thalli are found, i.e., thalli with chlorobionts growing on thalli with cyanobionts (James & Henssen, 1976). These observations were later confirmed by laboratory experiments and molecular data, and several additional pairs of cyanomorphs and chloromorphs were detected and described (e.g., Armaleo & Clerc, 1991; Galloway, 2001; James & Henssen, 1976; Magain et al., 2012; Moncada, Coca, & Lücking, 2013; Purvis, 2000; Ranft et al., 2018; Stocker-Wörgötter, 2002). Moreover, different photobionts can be recruited when the environment changes due to natural disasters, e.g., falling of trees to form open habitats. At a few sites in Bolivia, thalli with cyanobacteria were deteriorating and being overgrown by the attached thalli with green algae (Kukwa, unpublished; Figure 1A).

Shift in photobiont type may alter the morphology, such as color of the thallus or tomentum (Moncada, Coca, & Lücking, 2013) as well as thallus organization. Some species, when associated with cyanobacteria, form fruticose and branched thalli, which have been placed in the genus *Dendriscocaulon* (Galloway, 2001; James & Henssen, 1976; Magain et al., 2012; Purvis, 2000; Ranft et al., 2018; Tønsberg & Goward, 2001). Interestingly, however, *Dendriscocaulon*-like cyanomorphs of the genus *Sticta* are mostly found in temperate regions, whereas thalli with cyanobacteria are foliose and more similar in morphology to their green algal counterparts in the tropics (Moncada, Coca, & Lücking, 2013).

In a very few cases (e.g., *Sticta phyllidiokunthii* Moncada & Lücking), the switch of photobionts also alters the mode of reproduction, as cyanomorphs develop vegetative propagules in addition to apothecia, whereas chloromorphs develop apothecia alone (James & Henssen, 1976; Moncada, Coca, & Lücking, 2013).

#### 3. Dendriscocaulon - A Heterogenic Assemblage of Species

*Dendriscocaulon*, introduced by Nylander (1885) for *D. bolacinum* (Ach.) Nyl., accommodates species with branched thalli and cyanobionts. Numerous

species have been described, but several studies have also reported unnamed Dendriscocaulon-like thalli (often as cephalodia) associated with lichens containing green algae as the main photobiont (e.g., Clerc et al., 1992; Galloway, 1983, 2001; James & Henssen, 1976; Magain et al., 2012; Tønsberg & Goward, 2001). The association between both photomorphs was speculated for years, although in some cases, it was difficult to prove that they represented the same species when cyanomorph and chloromorph were allopatric (James & Henssen, 1976). Thanks to molecular techniques, this problem could be solved. Sequencing of fungal molecular markers showed that Dendriscocaulon-like lichens are, in fact, cyanomorphs of species accommodated in the genera Dendriscosticta Moncada & Lücking, Sticta, and Ricasolia (Figure 1B) and that the genus itself is a heterogeneous assemblage of distantly related species (e.g., Galloway, 2001; Magain et al., 2012; Moncada, Coca, & Lücking, 2013; Moncada, Lücking, & Betancourt-Macuase, 2013; Ranft et al., 2018; Stenroos et al., 2003). Therefore, formally described Dendriscocaulon species have now become synonyms of other species and the generic name itself is a synonym of Ricasolia (Galloway, 2001; Moncada, Lücking, & Betancourt-Macuase, 2013).

Symbiosis with two different photobionts may also have chemical consequences. James and Henssen (1976) studied 100 free-living *Dendriscocaulon*-like thalli using thin-layer chromatography or microcrystal tests and found no lichen secondary metabolites. However, the chloromorph may produce a few lichen substances, e.g., scrobiculin present in the chloromorphs of *Ricasolia amplissima* (Scop.) De Not. s. l. (for the most recent taxonomy of the species, see Cornejo et al., 2017). The nature of this phenomenon is unknown, but perhaps, the presence of cyanobacteria actively inhibits the production of certain lichen substances (James & Henssen, 1976). Meanwhile, this type of photobiont also alters the products of nitrogen metabolism: Ammonia and amines are produced by cyanomorphs and, when the thallus is wet, are released, emitting fish-like odor (James & Henssen, 1976; see also previous section).

# 4. *Buellia violacefusca* and *Lecanographa amylacea* – When Two Become One

A single lichen thallus of the same species can contain numerous species or OTUs of closely related photobionts, and different thalli may also be associated with different phylogenetic lineages of photobionts of the same genus (e.g., del Campo et al., 2013; Dal Grande et al., 2017; Friedl, 1987; Guzow-Krzemińska, 2006; Moya et al., 2017; Muggia et al., 2014; Nelsen & Gargas, 2008; Onut-Brännström et al., 2018). Occasionally, although a lichen recruits photobionts of two closely related green algal genera (Engelen et al., 2010), there are no changes in morphology, reproductive methods, or secondary metabolites. However, the shift in the symbiont from the green algal genus *Trebouxia* in the class Trebouxiophyceae Friedl to *Trentepohlia* s. l. in the class Ulvophyceae K. R. Mattox & K. D. Stewart may alter the reproductive mode and phenotypic dimorphism (Ertz et al., 2018).

*Buellia violaceofusca* G. Thor & Muhr was described as a sterile, sorediate lichen with blue-brown soralia and green algae as the photobiont. Due to the absence of apothecia, its taxonomic position was unclear, but owing to morphological similarity to *Buellia griseovirens* (Turner & Borrer ex Sm.), it was tentatively placed in the genus *Buellia* De Not. (Thor & Muhr, 1991) in the class Lecanoromycetes O. E. Erikss. & Winka (Wijayawardene et al., 2020). When fresh material of the species became available, it was sequenced. Surprisingly, analyses of fungal mtSSU and ITS rDNA markers placed it within the genus *Lecanographa* Egea & Torrente in the class Arthoniomycetes O. E. Erikss. & Winka (Ertz et al., 2018). Sequences appeared to be identical to *Lecanographa amylacea* (Ehrh. ex Pers.) Egea & Torrente, which lacks soredia, but reproduces sexually, forming thalli with a trentepohlioid photobiont that contains large amounts of carotenoid pigments, causing the algae to appear yellow-orange (Ertz et al., 2018). *Buellia violaceofusca* and *Lecanographa amylacea* are conspecific, the latter being the oldest available name for this lichenized fungus (Ertz et al., 2018).

This is a well-documented example of a lichen species recruiting two different green algae of distantly related genera. A switch between these unrelated photosynthetic

partners is obviously responsible for thallus dimorphism and has a major impact on the anatomy, morphology, and reproductive strategy of the species. The *Trebouxia*morphotype of *Lecanographa amylacea* is always sterile and sorediate, contrary to its trentepohlioid morphotype, which never produces soredia. When *Lecanographa amylacea* is fertile, its ascospores can capture *Trebouxia* algae from other lichens (or perhaps also from free-living, nonlichenized *Trebouxia*) to form the sorediate thallus, but when trentepohlioid algae are recruited, it develops esorediate, fertile thalli. This flexibility can be considered as a strategy to increase habitat tolerance, which allows the lichen to withstand environmental changes. Examination of numerous specimens revealed that the sorediate component developed when the esorediate morphotype of *Lecanographa amylacea* grew in proximity to *Chrysothrix candelaris* (L.) J. R. Laundon; soredia developed in the areas where hyphae of the former invaded the thallus of the latter. Molecular data showed that both species often share the same *Trebouxia* strain (Ertz et al., 2018).

*Lecanographa amylacea* is a rare lichen, but its sorediate morphotype is more common and reported from stands where fertile thalli do not grow (e.g., Ertz et al., 2018; Kukwa et al., 2012; Poelt, 1994; Thor & Muhr, 1991). This observation indicates that the strategy to recruit two different photobionts and the shift of the reproductive mode can be evolutionarily advantageous.

This is the first and thus far the only documented case of a single lichen species recruiting *Trebouxia* and *Trentepohlia* s. l. photobionts, which have altered its morphology and reproductive mode. However, by studying sterile sorediate or isidiate lichens of the family Graphidaceae in Bolivia, we found that this phenomenon may exist in a few cases within this group of lichenized fungi. Most members of Graphidaceae form thalli with trentepohlioid photobionts (e.g., Kosecka et al., 2020; Rivas Plata et al., 2010; Staiger, 2002), but in some sterile Bolivian samples of this group, Trebouxiophyceae photobionts were detected using morphological and molecular approaches. However, their fertile photomorph with *Trentepholia*-like algae have not been found yet. More details will be presented in a forthcoming article.

# 5. *Ionaspis/Hymenelia complex* – Is the Photobiont Not Enough to Distinguish Genera?

The genera Ionaspis Th. Fr. and Hymenelia Kremp. (Hymeneliaceae Körb.) were traditionally separated on the basis of their different photobionts, trentepohlioid or trebouxioid, respectively. However, Eigler (1969) showed heterogeneity within this complex, with some *Ionaspis* species being more similar to *Hymenelia* species than to their congeners. Therefore, the distinction of these genera based on the photobiont was questioned by numerous lichenologists (e.g., Eigler, 1969; Jørgensen, 1989; Lutzoni & Brodo, 1995). By employing cladistic analyses of morphological, anatomical, and allozyme data, Lutzoni and Brodo (1995) reviewed North American taxa within the Ionaspis/Hymenelia complex and reclassified some species. They treated the photobiont as a minor character and separated these two genera on the basis of differences in epihymenial pigments, ascospore width, and hymenium thickness (Lutzoni & Brodo, 1995). Although their new classification is generally accepted, the distinction between these two genera was claimed to be practically difficult and artificial compared to the photobiont-based classification, and the need for morphological and molecular revisions of these taxa was thus highlighted (Fryday & McCarthy, 2018; Kantvilas, 2014). However, this complex has not been studied in more detail using DNA data, with the exception of a study based on limited sampling from Alaska, which showed that this group of lichens warrants further investigation (McCune et al., 2018).

#### 6. Stictidaceae and Optional Lichenization

The family Stictidaceae Fr. comprises 28 genera (Wijayawardene et al., 2020) of lichenized, lichenicolous, and saprotrophic fungi (e.g., Diederich et al., 2018; Wedin et al., 2004, 2005, 2006). This also includes the genus *Stictis* Pers., which originally comprised epixylic, nonlichenized saprotrophic fungi (Wedin et

al., 2004, 2005, 2006). However, molecular data also placed species of the former *Conotrema* Tuck., which are lichenized and epiphytic fungi, within the genus *Stictis* (Wedin et al., 2004, 2005, 2006). Although such a situation is not an exception in fungi, it is remarkable in this case. Using molecular markers, Wedin et al. (2004, 2005, 2006) discovered that neither the genera nor the species are monophyletic. Moreover, the same species, depending on the substrate (bark or wood), either formed a lichenized *Conotrema*-like thallus or lived as a saprotroph without symbiotic algae (*Stictis*-like thallus) and completed its lifecycle. The phenomenon that a single species can exist as a lichen or a saprotroph is called optional lichenization (Wedin et al., 2004, 2005, 2006).

Individuals of *Stictis* species that can form lichenized and nonlichenized thalli differ in their morphology. Ascomata of both forms are very similar, but as saprotrophs, they have generally more heavily pigmented ascomatal walls and are more exposed on the substrate. Additionally, individuals living in symbiosis with algae form highly visible, whitish, lichenized thalli on tree barks, whereas thalli of individuals lacking photobionts are less evident, completely immersed in the substratum and only their ascomata are visible. A very similar situation, with two nutritional modes (lichenized and saprotrophic), is also known in another member of Stictidaceae: *Schizoxylon albescens* Gilenstam, H. Döring & Wedin (Fernández-Brime et al., 2019; Wedin et al., 2006).

Optional lichenization may play important roles in the evolution of Stictidaceae, representing an advantageous adaptive strategy allowing the species to grow on different substrates. It enables the species to colonize tree barks (mostly *Populus tremula*) when there is no lignum available for the nonlichenized form in the habitat or when the lignum is already present but the host trees are too young for colonization of the lichenized form (Wedin et al., 2004). Moreover, species with two different nutritional modes may be more common and widespread than species lacking this potential (Wedin et al., 2004, 2006).

#### 7. Conclusions

The switch of photobionts or optional lichenization are the two potential evolutionarily advantageous strategies that allow lichens to occupy a wider range of habitats. Recruitment of different photobionts or optional lichenization allows the species to colonize habitats with diverse conditions of light and humidity (e.g., Ertz et al., 2018; Green et al., 1993; James & Henssen, 1976; Purvis, 2000) or occupy different substrates (Fernández-Brime et al., 2019; Wedin et al., 2004, 2005, 2006). Both these strategies alter the anatomy, morphology, secondary metabolites, and reproductive modes (Ertz et al., 2018; James & Henssen, 1976; Lutzoni & Brodo, 1995; Magain et al., 2012; Moncada, Coca, & Lücking, 2013; Purvis, 2000; Wedin et al., 2004) as well as influence the taxonomy and classification of lichenized fungi.

According to Turland et al. (2018), the taxonomy of lichens is always based on the fungal partner (Article F.1.1.); therefore, only a single name must be used for a fungus associated with different photobionts (or in the case of optional lichenization, lacking photobionts), despite variations in the morphology, chemical composition, and reproductive modes caused by association with different photobionts. However, before the application of molecular methods, the identification and classification of such different morphs of a single species were limited and often separate names were applied (e.g., Ertz et al., 2018; Heidmarsson et al., 1997; Jørgensen, 1998; Laundon, 1995; Tønsberg & Goward, 2001). Laundon (1995) proposed forma as a taxonomic rank to recognize photomorphs; however, Heidmarsson et al. (1997) claimed that it was illogical to use any taxonomic rank in such cases, particularly when two photomorphs can form an association in a single lichen thallus [like in Sticta canariensis (Bory) Bory ex Delise]. These authors also stated that any rank should reflect genetic differences - a point of view accepted in subsequent studies (e.g., Ertz et al., 2018; Miadlikowska et al., 2018; Moncada, Coca, & Lücking, 2013; Purvis, 2000; Ranft et al., 2018).

#### Acknowledgments

Dr. William Sanders is acknowledged for discussion on *Hymenelia*, Prof. Mark R. D. Seaward (Bradford) for advice and language improvement, and two reviewers for helpful comments.

#### References

- Armaleo, D., & Clerc, P. (1991). Lichen chimeras: DNA analysis suggests that one fungus forms two morphotypes. *Experimental Mycology*, 15, 1–10. https://doi.org/10.1016/0147-5975(91)90002-U
- Arnold, A. E., Miadlikowska, J., Higgins, K. L., Sarvate, S. D., Gugger, P., Way, A., Hofstetter, V., Kauff, F., & Lutzoni, F. (2009). A phylogenetic estimation of trophic transition networks for ascomycetous fungi: Are lichens cradles of symbiotrophic fungal diversification? *Systematic Biology*, 58(3), 283–297. https://doi.org/10.1093/sysbio/syp001
- Bates, S. T., Berg-Lyons, D., Lauber, C. L., Walters, W. A., Knight, R., & Fierer, N. (2012). A preliminary survey of lichen associated eukaryotes using pyrosequencing. *Lichenologist*, 44(1), 137–146. https://doi.org/10.1017/S0024282911000648
- Bates, S. T., Cropsey, G. W. G., Caporaso, J. G., Knight, R., & Fierer, N. (2011). Bacterial communities associated with the lichen symbiosis. *Applied and Environmental Microbiology*, 77(4), 1309–1314. https://doi.org/10.1128/AEM.02257-10
- Clerc, P., Camenzind, R., Dietrich, M., Groner, U., Grundlehner, S., Oberli, F., Scheidegger, C., & Wildi, E. (1992). *Lobaria amplissima* (Scop.) Forss. dans les Préalpes vaudoises [*Lobaria amplissima* (Scop.) Forss. in the Western Alps]. *Meylania*, 1(1), 16–20.
- Cornejo, C., Derr, C., & Dillman, K. (2017). *Ricasolia amplissima* (Lobariaceae): One species, three genotypes and a new taxon from south-eastern Alaska. *Lichenologist*, 49(6), 579–596. https://doi.org/10.1017/S002428291700041X
- Dal-Forno, M., Lawrey, J. D., Sikaroodi, M., Bhattarai, S., Gillevet, P. M., Sulzbacher, M., & Lücking, R. (2013). Starting from scratch: Evolution of the lichen thallus in the basidiolichen *Dictyonema* (Agaricales: Hygrophoraceae). *Fungal Biology*, 117(9), 584–598. https://doi.org/10.1016/j.funbio.2013.05.006
- Dal Grande, F., Rolshausen, G., Divakar, P. K., Crespo, A., Otte, J., Schleuning, M., & Schmitt, I. (2017). Environment and host identity structure communities of green algal symbionts in lichens. *New Phytologist*, *217*(1), 277–289. https://doi.org/10.1111/nph.14770
- del Campo, E., Catalá, S., Gimeno, J., del Hoyo, A., Martínez-Alberola, F., Casano, L. M., Grube, M., & Barreno, E. (2013). The genetic structure of the cosmopolitan threepartner lichen *Ramalina farinacea* evidences the concerted diversification of symbionts. *FEMS Microbiology Ecology*, *83*(2), 310–323. https://doi.org/10.1111/j.1574-6941.2012.01474.x
- Diederich, P., Lawrey, J. D., & Ertz, D. (2018). The 2018 classification and checklist of lichenicolous fungi, with 2000 non-lichenized, obligately lichenicolous taxa. *Bryologist*, 121(3), 340–425. https://doi.org/10.1639/0007-2745-121.3.340
- Eigler, G. (1969). *Studien zur Gliederung der Flechtengattung Lecanora* [Studies on the classification of the lichen genus *Lecanora*]. Lubrecht & Cramer.
- Engelen, A., Convey, P., & Ott, S. (2010). Life history strategy of *Lepraria borealis* at an Antarctic inland site, Coal Nunatak. *Lichenologist*, 42(3), 339–346. https://doi.org/10.1017/S0024282909990600
- Ertz, D., Guzow-Krzemińska, B., Thor, G., Łubek, A., & Kukwa, M. (2018). Photobiont switching causes changes in the reproduction strategy and phenotypic dimorphism in the Arthoniomycetes. *Scientific Reports*, *8*, Article 4952. https://doi.org/10.1038/s41598-018-23219-3
- Fernández-Brime, S., Muggia, L., Maier, S., Grube, M., & Wedin, M. (2019). Bacterial communities in an optional lichen symbiosis are determined by substrate, not algal photobionts. *FEMS Microbiology Ecology*, 95(3), Article fiz012. https://doi.org/10.1093/femsec/fiz012
- Fernández-Mendoza, F., Fleischhacker, A., Kopun, T., Grube, M., & Muggia, L. (2017). ITS1 metabarcoding highlight slow specificity of lichen mycobiomes at a local scale. *Molecular Ecology*, 26(18), 4811–4830. https://doi.org/10.1111/mec.14244
- Flakus, A., Etayo, J., Miadlikowska, J., Lutzoni, F., Kukwa, M., Matura, N., & Rodriguez-Flakus, P. (2019). Biodiversity assessment of ascomycetes inhabiting *Lobariella*

lichens in Andean cloud forests led to one new family, three new genera and 13 new species of lichenicolous fungi. *Plant and Fungal Systematics*, 64(2), 283–344. https://doi.org/10.2478/pfs-2019-0022

- Friedl, T. (1987). Thallus development and phycobionts of the parasitic lichen *Diploschistes* muscorum. Lichenologist, 19(2), 183–191. https://doi.org/10.1017/S002428298700015X
- Friedl, T., & Büdel, B. (2008). Photobionts. In T. H. Nash III (Ed.), *Lichen biology* (2nd ed., pp. 7–26). Cambridge University Press. https://doi.org/10.1017/CBO9780511790478.003
- Fryday, A. M., & McCarthy, J. W. (2018). Hymenelia parva (Hymeneliaceae, Ostropomycetidae): A new species from Newfoundland, Canada. Graphis Scripta, 30(6), 44–50.
- Galloway, D. J. (1983). New taxa in the New Zealand lichen flora. New Zealand Journal of Botany, 21, 191–200. https://doi.org/10.1080/0028825X.1983.10428544
- Galloway, D. J. (1994). Studies on the lichen genus *Sticta* (Schreber) Ach.: I. Southern South American species. *Lichenologist*, *26*(3), 223–282. https://doi.org/10.1006/lich.1994.1019
- Galloway, D. J. (1997). Studies on the lichen genus *Sticta* (Schreber) Ach. IV. New Zealand species. *Lichenologist*, *29*(2), 105–168. https://doi.org/10.1006/lich.1996.0066
- Galloway, D. J. (1998). Studies on the lichen genus *Sticta* (Schreber) Ach.: V. Australian species. *Tropical Bryology*, *15*, 117–160. https://doi.org/10.11646/bde.15.1.12
- Galloway, D. J. (2001). "Dendriscocaulon". In P. M. McCarthy (Ed.), Flora of Australia. Volume 58A. Lichens 3 (pp. 38–39). ABRS/CSIRO.
- Galloway, D. J., & Nash, T. H., III. (2004). Sticta. In T. H. Nash III, B. D. Ryan, P. Diederich, C. Gries, & F. Bungartz (Eds.), Lichen flora of the greater Sonoran Desert region (Vol. 2, pp. 513–524). Lichens Unlimited.
- Green, T. G. A., Büdel, B., Heber, U., Meyer, A., Zellner, H., & Lange, O. L. (1993). Differences in photosynthetic performance between cyanobacterial and green algal components of lichen photosymbiodemes measured in the field. *New Phytologist*, *125*(4), 723–731. https://doi.org/10.1111/j.1469-8137.1993.tb03921.x
- Grube, M., Cernava, T., Soh, J., Fuchs, S., Aschenbrenner, I., Lassek, C., Wegner, U., Becher, D., Riedel, K., Sensen, C. W., & Berg, G. (2015). Exploring functional contexts of symbiotic sustain within lichen-associated bacteria by comparative omics. *Multidisciplinary Journal of Microbial Ecology*, 9(2), 412–424. https://doi.org/10.1038/ismej.2014.138
- Guzow-Krzemińska, B. (2006). Photobiont flexibility in the lichen *Protoparmeliopsis muralis* as revealed by ITS rDNA analysis. *Lichenologist*, 38(5), 469–476. https://doi.org/10.1017/S0024282906005068
- Hawksworth, D. L., Santesson, R., & Tibell, L. (2011). *Racoleus*, a new genus of sterile filamentous lichen-forming fungi from the tropics, with observations on the nomenclature and typification of *Cystocoleus* and *Racodium*. *IMA Fungus*, 2(1), 71–79. https://doi.org/10.5598/imafungus.2011.02.01.10
- Heidmarsson, S., Mattsson, J. E., Moberg, R., Nordin, A., Santesson, R., & Tibell, L. (1997). Classification of lichen photomorphs. *Taxon*, 46(3), 519–520. https://doi.org/10.2307/1224392
- James, P. W., & Henssen, A. (1976). The morphological and taxonomic significance of cephalodia. In D. H. Brown, D. L. Hawksworth, & R. H. Bailey (Eds.), *Lichenology: Progress and problems* (pp. 27–77). Academic Press.
- Jørgensen, P. M. (1989). Notes on the lichen genus *Ionaspis* in Scandinavia. *Graphis Scripta*, 2(3), 118–121.
- Jørgensen, P. M. (1998). What shall we do with the blue-green counterparts? *Lichenologist*, 30(4–5), 351–356. https://doi.org/10.1006/lich.1998.0146
- Kantvilas, G. (2014). The lichen family Hymeneliaceae in Tasmania, with the description of a new species. *Kanunnah*, 7, 127–140.
- Kosecka, M., Jabłońska, A., Flakus, A., Rodriguez-Flakus, P., Kukwa, M., & Guzow-Krzemińska, B. (2020). Trentepohlialean algae (Trentepohliales, Ulvophyceae) show preference to selected mycobiont lineages in lichen symbioses. *Journal of Phycology*, 56(4), 979–993. https://doi.org/10.1111/jpy.12994
- Kukwa, M., Łubek, A., Szymczyk, R., & Zalewska, A. (2012). Seven lichen species new to Poland. *Mycotaxon*, 120, 105–118. https://doi.org/10.5248/120.105
- Lamb, I. M. (1951). On the morphology, phylogeny, and taxonomy of the lichen genus Stereocaulon. Canadian Journal of Botany, 29, 522–584. https://doi.org/10.1139/b51-045
- Laundon, J. R. (1995). On the classification of lichen photomorphs. *Taxon*, 44(3), 387–389. https://doi.org/10.2307/1223409

- Lutzoni, F. M., & Brodo, I. M. (1995). A generic redelimitation of the *Ionaspis– Hymenelia* complex (lichenized Ascomycotina). *Systematic Botany*, 20(3), 224–258. https://doi.org/10.2307/2419494
- Lücking, R., Dal Forno, M., Moncada, B., Coca, F. L., Vargas-Mendoza, L. J., Aptroot, A., Arias, L. J., Besal, B., Bungartz, F., Cabrera-Amaya, D. M., Cáceres, M. E. S., Chaves, J. F., Eliasaro, S., Gutiérrez, M. C., Marin, J. H., de los Ángeles Herrera-Campos, M., Holgado-Rojas, M. E., Jonitz, H., Kukwa, M., ... Lawrey, J. D. (2017). Turbo-taxonomy to assemble a megadiverse lichen genus: Seventy new species of *Cora* (Basidiomycota: Agaricales: Hygrophoraceae), honouring David Leslie Hawksworth's seventieth birthday. *Fungal Diversity*, 84, 139–207. https://doi.org/10.1007/s13225-016-0374-9
- Lücking, R., Hodkinson, B. P., & Leavitt, S. D. (2017). The 2016 classification of lichenized fungi in the Ascomycota and Basidiomycota – Approaching one thousand genera. *Bryologist*, 119(4), 361–416. https://doi.org/10.1639/0007-2745-119.4.361
- Lücking, R., Lawrey, J. D., Sikaroodi, M., Gillevet, P. M., Chaves, J. L., Sipman, H. J. M., & Bungartz, F. (2009). Do lichens domesticate photobionts like farmers domesticate crops? Evidence from a previously unrecognized lineage of filamentous cyanobacteria. *American Journal of Botany*, 96(8), 1409–1418. https://doi.org/10.3732/ajb.0800258
- Magain, N., Goffinet, B., & Sérusiaux, E. (2012). Further photomorphs in the lichen family Lobariaceae from Reunion (Mascarene archipelago) with notes on the phylogeny of *Dendriscocaulon cyanomorphs. Bryologist*, 115(2), 243–254. https://doi.org/10.1639/0007-2745-115.2.243
- McCune, B., Arup, U., Breuss, O., Di Meglio, E., Di Meglio, J., Esslinger, T. L., Magain, N., Miadlikowska, J., Miller, A. E., Muggia, L., Nelson, P. R., Rosentreter, R., Schultz, M., Sheard, J. W., Tønsberg, T., & Walton, J. (2018). Biodiversity and ecology of lichens of Katmai and Lake Clark national parks and preserves, Alaska. *Mycosphere*, 9(4), 859–930. https://doi.org/10.5943/mycosphere/9/4/10
- Miadlikowska, J., Kauff, F., Hofstetter, V., Fraker, E., Grube, M., Hafellner, J., Reeb, V., Hodkinson, B. P., Kukwa, M., Lücking, R., Hestmark, G., Otalora, M. G., Rauhut, A., Büdel, B., Scheidegger, C., Timdal, E., Stenroos, S., Brodo, I., Perlmutter, G. B., ... Lutzoni, F. (2006). New insights into classification and evolution of the Lecanoromycetes (Pezizomycotina, Ascomycota) from phylogenetic analyses of three ribosomal RNA- and two protein-coding genes. *Mycologia*, *98*(6), 1088–1103. https://doi.org/10.3852/mycologia.98.6.1088
- Miadlikowska, J., Magain, N., Pardo-De la Hoz, C. J., Niu, D., Goward, T., Sérusiaux, E., & Lutzoni, F. (2018). Species in section *Peltidea (aphthosa* group) of the genus *Peltigera* remain cryptic after molecular phylogenetic revision. *Plant and Fungal Systematics*, 63(2), 45–64. https://doi.org/10.2478/pfs-2018-0007
- Moncada, B., Coca, L. F., & Lücking, R. (2013). Neotropical members of *Sticta* (lichenized Ascomycota: Lobariaceae) forming photosymbiodemes, with the description of seven new species. *Bryologist*, 116(2), 169–200. https://doi.org/10.1639/0007-2745-116.2.169
- Moncada, B., Lücking, R., & Betancourt-Macuase, L. (2013). Phylogeny of the Lobariaceae (lichenized Ascomycota: Peltigerales), with a reappraisal of the genus *Lobariella*. *Lichenologist*, 45(2), 203–263. http://doi.org/10.1017/S0024282912000825
- Moya, P., Molins, A., Martinez-Alberola, F., Muggia, L., & Barreno, E. (2017). Unexpected associated microalgal diversity in the lichen *Ramalina farinacea* is uncovered by pyrosequencing analyses. *PLoS One*, *12*(4), Article e0175091. https://doi.org/10.1371/journal.pone.0175091
- Muggia, L., Fleischhacker, A., Kopun, T., & Grube, M. (2016). Extremotolerant fungi from alpine rock lichens and their phylogenetic relationships. *Fungal Diversity*, *76*, 119–142. https://doi.org/10.1007/s13225-015-0343-8
- Muggia, L., Pérez-Ortega, S., Kopun, T., Zellnig, G., & Grube, M. (2014). Photobiont selectivity leads to ecological tolerance and evolutionary divergence in a polymorphic complex of lichenized fungi. *Annals of Botany*, *114*(3), 463–475. https://doi.org/10.1093/aob/mcu146
- Nelsen, M. P., & Gargas, A. (2008). Dissociation and horizontal transmission of codispersing lichen symbionts in the genus *Lepraria* (Lecanorales: Stereocaulaceae). *New Phytologist*, 177(1), 264–275. https://doi.org/10.1111/j.1469-8137.2007.02241.x
- Nylander, W. (1885). Addenda nova ad Lichenographiam Europaeam. Continuatio quadragesima quarta [A new addition to the European lichens]. *Flora*, 68, 295–301.
- Onuţ-Brännström, I., Benjamin, M., Scofield, D. G., Heiðmarsson, S., Andersson, M. G. I., Lindström, E. S., & Johannesson, H. (2018). Sharing of photobionts in sympatric populations of *Thamnolia* and *Cetraria* lichens: Evidence from high-throughput sequencing. *Scientific Reports*, 8, Article 4406. https://doi.org/10.1038/s41598-018-22470-y

- Oset, M., & Kukwa, M. (2012). Notes on *Stereocaulon* species from Bolivia. *Mycotaxon*, 121, 447–453. https://doi.org/10.5248/121.447
- Poelt, J. (1994). Bemerkenswerte Flechten aus Österreich, insbesondere der Steiermark [Remarkable collections of lichens from Austria, especially from Styria]. *Mitteilungen der Naturwissenschaftlichen Vereines fur Steiermark*, 124, 91–111.
- Purvis, W. (2000). Lichens. Natural History Museum.
- Ranft, H., Moncada, B., Lange, P. J., Lumbsch, H. T., & Lücking, R. (2018). The *Sticta filix* morphodeme (Ascomycota: Lobariaceae) in New Zealand with the newly recognized species *S. dendroides* and *S. menziesii*: Indicators of forest health in a threatened island biota? *Lichenologist*, 50(2), 185–210. https://doi.org/10.1017/S0024282917000706
- Rivas Plata, E., Lücking, R., Sipman, H. J. M., Mangold, A., Kalb, K., & Lumbsch, H. T. (2010). A world-wide key to the thelotremoid Graphidaceae, excluding the *Ocellularia-Myriotrema-Stegobolus* clade. *Lichenologist*, 42(2), 139–185. https://doi.org/10.1017/S0024282909990491
- Schneider, K., Resl, P., & Spribille, T. (2016). Escape from the cryptic species trap: Lichen evolution on both sides of a cyanobacterial acquisition event. *Molecular Ecology*, 25(14), 3453–3468. https://doi.org/10.1111/mec.13636
- Singh, G., Kukwa, M., Dal Grande, F., Łubek, A., Otte, J., & Schmitt, I. (2019). A glimpse into genetic diversity and symbiont interaction patterns in lichen communities from areas with different disturbance histories in Białowieża Forest, Poland. *Microorganisms*, 7, Article 335. https://doi.org/10.3390/microorganisms7090335
- Spribille, T., Tuovinen, V., Resl, P., Vanderpool, D., Wolinski, H., Aime, M. C., Schneider, K., Stabentheiner, E., Toome-Heller, M., Thor, G., Mayrhofer, H., Johannesson, H., & McCutcheon, J. P. (2016). Basidiomycete yeasts in the cortex of ascomycete macrolichens. *Science*, 353(6298), 488–492. https://doi.org/10.1126/science.aaf8287
- Staiger, B. (2002). Die Flechtenfamilie Graphidaceae. Studien in Richtung einer natürlicheren Gliederung [The lichen family Graphidaceae. Studies towards a more natural systematics]. J. Cramer.
- Stenroos, S., Stocker-Wörgötter, E., Yoshimura, I., Myllys, L., Thell, A., & Hyvonen, J. (2003). Culture experiments and DNA sequence data confirm the identity of *Lobaria* photomorphs. *Canadian Journal of Botany*, 81(3), 232–247. https://doi.org/10.1139/b03-027
- Stocker-Wörgötter, E. (2002). Resynthesis of photosymbiodemes. In I. Kranner, R. P. Beckett, & A. K. Varma (Eds.), Protocols in lichenology. Culturing, biochemistry, ecophysiology and use in biomonitoring (pp. 47–60). Springer. https://doi.org/10.1007/978-3-642-56359-1\_3
- Thor, G., & Muhr, L. E. (1991). *Buellia violaceofusca*, a new lichen from Sweden. *Lichenologist*, 23(1), 11–13. https://doi.org/10.1017/S0024282991000051
- Tønsberg, T., & Goward, T. (2001). *Sticta oroborealis* sp. nov. and other Pacific North American lichens forming dendriscocauloid cyanotypes. *Bryologist*, *104*(1), 12–23. https://doi.org/d5gvbs
- Tschermak-Woess, E. (1988). The algal partner. In M. Galun (Ed.), *Handbook of lichenology* (Vol. 1, pp. 75–104). CRC Press.
- Tschermak-Woess, E. (1989). Developmental studies in trebouxioid algae and taxonomical consequences. *Plant Systematics and Evolution*, *164*, 161–195. https://doi.org/10.1007/BF00940436
- Turland, N. J., et al. (Eds.). (2018). International Code of Nomenclature for algae, fungi, and plants (Shenzhen Code) adopted by the Nineteenth International Botanical Congress Shenzhen, China, July 2017. Koeltz Botanical Books. https://doi.org/10.12705/Code.2018
- Wedin, M., Döring, H., & Gilenstam, G. (2004). Saprotrophy and lichenization as options for the same fungal species on different substrata: Environmental plasticity and fungal lifestyles in the *Stictis-Conotrema* complex. *New Phytologist*, *164*(3), 459–465. https://doi.org/10.1111/j.1469-8137.2004.01198.x
- Wedin, M., Döring, H., & Gilsenstam, G. (2006). *Stictis* s. lat. (Ostropales, Ascomycotina) in northern Scandinavia, with a key and notes on morphological variation in relation to lifestyle. *Mycological Research*, 110(7), 773–789. https://doi.org/10.1016/j.mycres.2006.04.010
- Wedin, M., Döring, H., Könberg, K., & Gilenstam, G. (2005). Generic delimitations in the family Stictidaceae (Ostropales, Ascomycota): The *Stictis–Conotrema* problem. *Lichenologist*, 37(1), 67–75. https://doi.org/10.1017/S0024282904014653
- West, N. J., Parrot, D., Fayet, C., Grube, M., Tomasi, S., & Suzuki, M. T. (2017). Marine cyanolichens from different littoral zones are associated with distinct bacterial communities. *PeerJ*, 2018(7), Article 5208. https://doi.org/10.7717/peerj.5208

Wijayawardene, N. N., Hyde, K. D., Al-Ani, L. K. T., Tedersoo, L., Haelewaters, D.,
Rajeshkumar, K. C., Zhao, R. L., Aptroot, A., Leontyev, D. V., Saxena, R. K.,
Tokarev, Y. S., Dai, D. Q., Letcher, P. M., Stephenson, S. L., Ertz, D., Lumbsch, H. T.,
Kukwa, M., Issi, I. V., Madrid, H., ... Thines, M. (2020). Outline of fungi and fungi-like
taxa. *Mycosphere*, *11*(1), 1060–1456. https://doi.org/10.5943/mycosphere/11/1/8